

FUNDAMENTAL STUDY OF BIOACTIVE PEPTIDES
ON SELECTED *Alexandrium spp.* AT DIFFERENT
GROWTH PHASES

BY

NURUL ASHIMA BT HAMDAN

A thesis submitted in fulfillment of the requirement for the
degree of Master in Science (Biotechnology)

Kulliyyah of Science
International Islamic University Malaysia

JANUARY 2020

ABSTRACT

The naturally occurring phenomenon of harmful algae blooming (HAB) at the water column brought detrimental effects to the economy as well as the environmental health of the water ecosystem. Most cases of HABs reported in Malaysia waters are dominated by dinoflagellates *Alexandrium spp.* with some of the species were reported to cause shellfish poisoning. In this study, two *Alexandrium spp.*; *A. tamiyavanichii* and *A. leei* with different toxicity levels were analyzed using proteomic approaches involving two-dimensional PAGE and HPLC analysis. The growth pattern of both species were identified and compared by using protein profiles at each growth phases. Protein expression reduced throughout the growth phases of *A. tamiyavanichii* but elevated during stationary phase of *A. leei*. A short duration of stationary phase suggests the continuous expression of growth proteins in *A. leei*. GNAT family acetyltransferase and lipases were successfully identified enzyme protein in *A. tamiyavanichii* and *A. leei* respectively with growth regulatory functions. The toxin profiles of both species exhibited higher level of toxin content in *A. tamiyavanichii* with 88 mol % of total toxins recorded as compared to 12 mol% in *A. leei*. The highest toxin content was recorded during exponential phase of *A. tamiyavanichii* with dominance of GTX4 and STX congeners. Further investigation on the toxicity effects of *Alexandrium spp.* was studied by using oysters where the shellfish are fed with toxic *A. tamiyavanichii* and non-toxic *A. leei*. LC-MS results of the two up-regulated spots in oysters fed with *A. tamiyavanichii* were identified as arginine kinase and transgelin. The up-regulation of the proteins was induced as responsive mechanism towards accumulation of the toxic species *A. tamiyavanichii*. These responses reflect the oyster stability during HAB outbreak. Fundamental studies of dinoflagellates from its molecular as well as byproduct analysis are useful to understand the biochemistry of the HAB species. The findings from this study can provide the basis knowledge on the biochemical properties of HAB species and the behavioral of affected organisms.

خلاصة البحث

لقد سببت ظاهرة انتشار الطحالب المضرة في الماء آثارا مدمرة للاقتصاد وللصحة البيئية والنظام البيئي المائي. أن معظم حالات انتشار الطحالب المضرة الموثقة في المياه الماليزية يسيطر عليها رتبة الأليكساندريوم من شعبة السوطيات الدوارة، حيث تسبب بعض الأنواع تسمم الصدفيات. تم في هذه الدراسة تحليل نوعي أليكساندريوم تاميافانيتشي (*Alexandrium tamiyavanichii*) وأليكساندريوم لبي (*Alexandrium leei*) بمستويات سمية مختلفة باستخدام الفصل الكهربائي لهلام كبريتات دوديكل الصوديوم متعدد الأكريلاميد (PAGE) والتحليل الكروماتوغرافي السائل العالي الأداء (HPLC). تم تحديد نمط النمو لكلا النوعين وتحليلهما بشكل أوسع عن طريق استخدام التنميط البروتيني في كل مراحل النمو. تم تخفيض التعبير البروتيني طوال مراحل النمو لنوع أليكساندريوم تاميافانيتشي، ولكنه ارتقى خلال المرحلة الثابتة لنوع أليكساندريوم لبي. تم التعرف بنجاح على أسيتيل ترانسفيراز من عائلة GNAT في نوع أليكساندريوم تاميافانيتشي، والليازات في نوع أليكساندريوم لبي كبروتينات إنزيمية بوظائف تنظيم النمو. أظهرت بروفيلات السمية لكل من النوعين مستوى أعلى للسمية في نوع أليكساندريوم تاميافانيتشي بقيمة 88 مول نسبي من إجمالي السموم المسجلة، مقارنة بـ 12 مول نسبي للأليكساندريوم لبي. تم تسجيل أعلى محتوى للسموم خلال طور النمو المطرد لأليكساندريوم تاميافانيتشي مع هيمنة مجانسات GTX4 و STX. تم القيام بالمزيد من التحقيقات في آثار سمية رتبة الأليكساندريوم في المحار حيث تم تغذية المحار بالأليكساندريوم تاميافانيتشي السام وأليكساندريوم لبي غير السام. تم تحديد نتائج LC-MS من المناطق المرفوعة في المحارات التي تغذيها بأليكساندريوم تاميافانيتشي بكونها كينازات الأرجينين وترانسجيلين. تم استنتاج أن التنظيم الرفعي للبروتينات كان آلية استجابة لتراكم النوع السام أليكساندريوم تاميافانيتشي في المحار. أظهر هذه الاستجابات قدرة تحمل المحار في مواجهة انتشار الطحالب المضرة. تعتبر الدراسات الأساسية لشعبة السوطيات الدوارة كتعبير جزئي وتحليل منتجاتها الثانوية مفيدة لفهم النشاط البيوكيميائي لأنواع الطحالب المضرة المنتشرة. بإمكان نتائج هذه الدراسة أن توفر معلومات أساسية حول الخصائص البيوكيميائية لأنواع الطحالب المضرة المنتشرة في المياه الماليزية وتأثيراتها المتسلسلة على النظام البيئي المائي.

APPROVAL PAGE

I certify that I have supervised and read this study and that in my opinion, it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master of Science (Biotechnology).

.....
Noraslinda Muhamad Bunnori
Supervisor

.....
Normawaty Mohammad Noor
Co-Supervisor

I certify that I have read this study and that in my opinion it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master of Science (Biotechnology).

.....
Noor Hasniza bt Md Zin
Internal Examiner

.....
Saiful Anuar bin Karsani
External Examiner

This thesis was submitted to the Department of Biotechnology and is accepted as a fulfillment of the requirements for the degree of Master of Science (Biotechnology).

.....
Mardiana Mohd Ashaari
Head, Department of Biotechnology

This thesis was submitted to the Kulliyyah of Science and is accepted as a fulfillment of the requirements for the degree of Master of Science (Biotechnology).

.....
Shahbudin bin Saad
Dean, Kulliyyah of Science

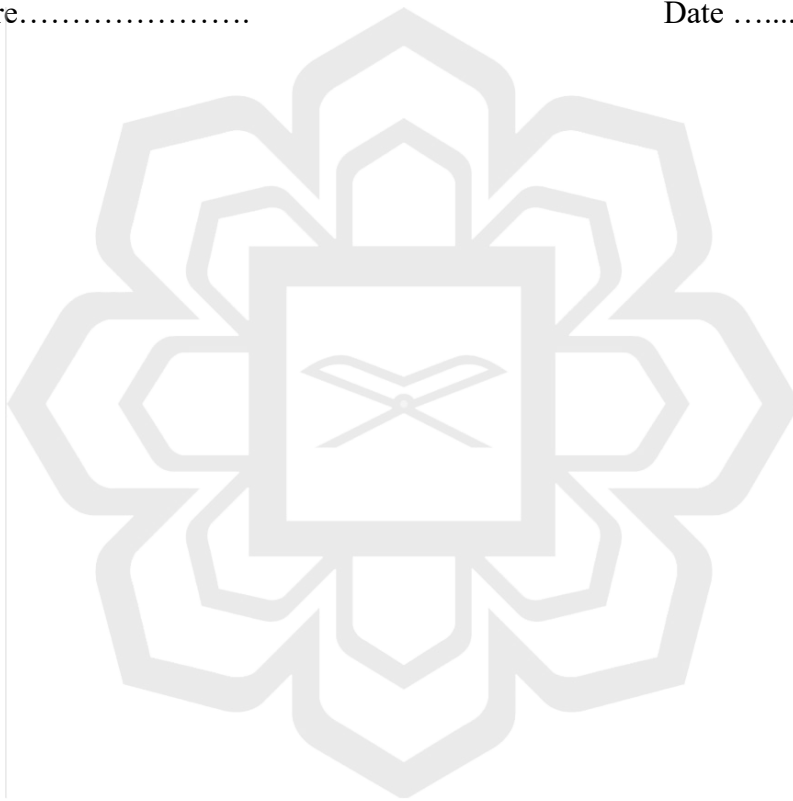
DECLARATION

I hereby declare that this thesis is the result of my own investigation, except where otherwise stated. I also declare that it has not been previously or concurrently submitted as a whole for any other degrees at IIUM or other institutions.

Nurul Ashima bt Hamdan

Signature.....

Date



INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

**DECLARATION OF COPYRIGHT AND AFFIRMATION OF
FAIR USE OF UNPUBLISHED RESEARCH**

**FUNDAMENTAL STUDY OF BIOACTIVE PEPTIDES ON
SELECTED *Alexandrium spp.* AT DIFFERENT
GROWTH PHASES**

I declare that the copyright holders of this thesis are jointly owned by the student and IIUM

Copyright © 2019 Nurul Ashima binti Hamdan International Islamic University Malaysia.
All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below.

1. Any material contained in or derived from this unpublished research may only be used by others in their writing with due acknowledgement.
2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purpose.
3. The IIUM library will have the right to make, store in a retrieval system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Nurul Ashima Hamdan

.....
Signature

.....
Date

ACKNOWLEDGEMENTS

Praise be to Allah, the Almighty and All-Knowing, for I could not have done this without His Mercy and Compassion. I would like to express my sincere gratitude to my supervisor Assistant Professor Dr. Noraslinda Muhamad Bunnori for all her valuable supervisory and advices in the field of biochemistry. I would also like to thank Associate Professor Dr. Normawaty bt Mohammad Noor for her helpful discussions in the field of phycology.

The work presented in this thesis has been performed at the Proteomic Laboratory of UIAM and Toxicology Laboratory of Kuantan Biosecurity in the time period 2015 to 2017. Special dedications to Sr. Suhaila and Sr. Hidayah for their assistance during my laboratory work. A warm gratitude to the science officers of Biosecurity, En. Bakri and En. Azlan for their cooperation throughout my toxicological study.

I am also grateful to all my colleagues and also the final year students for their help and idea throughout my study. My sincere thanks go to Bro Muhammad Shafiq for his continuous support upon completion of my thesis.

Finally I would like to thank my family for their love and understanding during these years. This is a gift for all of us who have contributed to its completion.

TABLE OF CONTENTS

Abstract	ii
Abstract in Arabic	iii
Approval page	iv
Declaration	v
Copyright Page.....	vi
Acknowledgement	vii
Table of Contents	viii
List of Tables	x
List of Figures	xi
List of Abbreviations	xiii
List of Symbols	xv
 CHAPTER ONE : BACKGROUND STUDY	 1
1.1 Harmful Algal Bloom (HAB)	1
1.2 Catastrophic Outcomes of HAB	2
1.2.1 Enormous Cases of Fish killing	2
1.2.2 Human Health Deterioration.....	3
1.3 Factors Influencing the Ovegrowth Cells	4
1.3.1 Nutrients Enrichment	5
1.3.2 Physical Oceanographic Changes	5
1.3.3 Environmental Parameters	6
1.4 <i>Alexandrium spp.</i> as Prominent Causative of HAB in Malaysia.....	7
1.5 Research Problem.....	8
1.6 Research Objectives.....	8
 CHAPTER TWO : PROTEIN PROFILING AT DIFFERENT	
GROWTH PHASES	9
2.1 Introduction.....	9
2.1.1 Proteomics of Dinoflagellates	9
2.1.2 Growth Phases of <i>Alexandrium spp</i>	9
2.1.3 Proteins Related to Overgrowth Cells.....	11
2.1.4 Toxin-related Proteins Produced during Blooming	12
2.2 Methodology	13
2.2.1 Growing of <i>Alexandrium spp</i>	13
2.2.2 Protein Extraction	14
2.2.3 Protein Quantification.....	15
2.2.4 One-dimensional Protein Separation by SDS-PAGE.. ..	16
2.2.5 Two-dimensional Protein Separation Performed using IEF	17
2.2.6 PDQuest Analysis.. ..	18
2.2.7 MALDI-TOF Analysis for Protein Identification.....	19
2.3 Results.....	20
2.3.1 Growth Curve of <i>A. tamiyavanichii</i> and <i>A. leei</i>	20
2.3.2 Developed Lysis Technique.....	22
2.3.3 Protein Concentrations.....	24
2.3.4. One-Dimensional Images.....	24

2.3.5 Two-Dimensional Images.....	26
2.4 Discussions	30
2.4.1 Growth Curve.....	30
2.4.2 Optimization of Lysis Technique.....	31
2.4.3 Protein Analysis	32
2.4.4 SDS-PAGE Analysis.....	33
2.4.5 Protein Identification.....	34
2.5 Conclusion	37
CHAPTER THREE: TOXIN PROFILING AT DIFFERENT GROWTH PHASES	38
3.1 Introduction.....	38
3.1.1 Toxin-producing <i>Alexandrium spp.</i>	38
3.1.2 Nutrients Inducing Toxicity	40
3.2 Methodology.....	41
3.2.1 Toxin Extraction	41
3.2.2 Toxins Analysis using HPLC.....	41
3.3 Results and Discussion	42
3.3.1 Toxicity of <i>A. tamiyavanichii</i> and <i>A. leei</i>	42
3.4 Conclusion	51
CHAPTER FOUR : PROTEIN EXPRESSION OF OYSTERS FED WITH DIFFERENT <i>Alexandrium spp.</i>.....	52
4.1 Introduction.....	52
4.1.1 Oysters as Filter Feeders	52
4.1.2 Response of Shellfish towards HAB.....	53
4.1.3 Proteins in Shellfish	55
4.2 Methodology.....	55
4.2.1 Oyster Acclimatization and Exposure Treatment	55
4.2.2 Protein Extraction	56
4.2.3 Protein Quantification.....	56
4.2.4 One-dimensional Protein Separation by SDS-PAGE	57
4.2.5 Two-dimensional Protein Separation.....	57
4.3 Results and Discussion	59
4.3.1 Protein Concentrations.....	59
4.3.2 One Dimensional Gel Electrophoresis.....	59
4.3.3 Two Dimensional Gel Electrophoresis	61
4.4 Conclusion	66
CHAPTER FIVE : CONCLUSION AND FUTURE DIRECTIONS.....	67
5.1 Conclusion	67
5.2 Future Directions	68
REFERENCES.....	69
APPENDIX.....	85

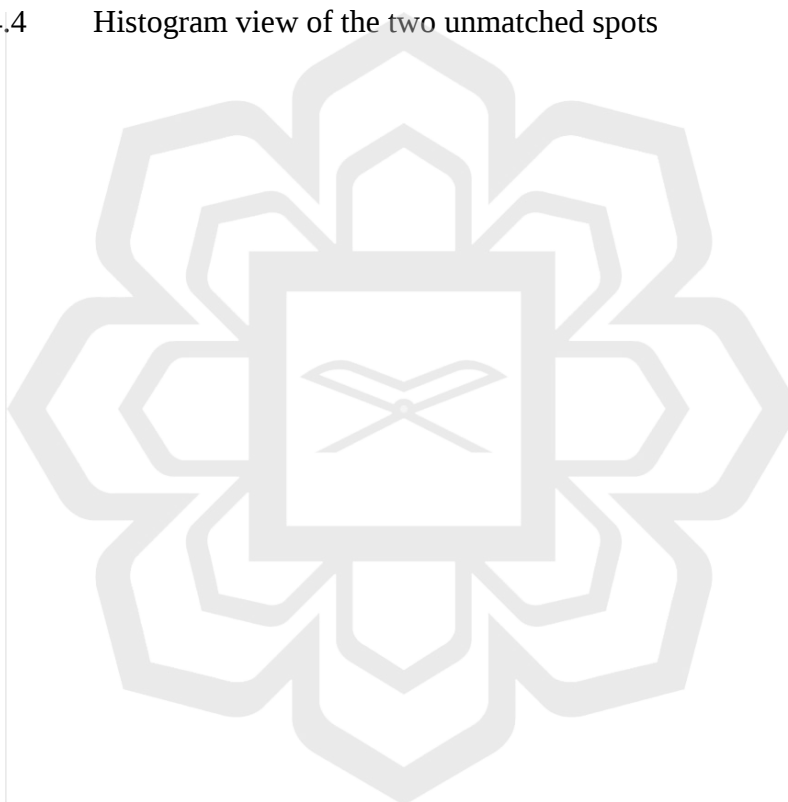
LIST OF TABLES

<u>Table No.</u>		<u>Page No.</u>
Table 2.1	Isoelectric focusing steps for 3-10 linear IPG strips	18
Table 2.2	Protein contents from different lysis techniques	22
Table 2.3	Protein contents from all growth phases of <i>Alexandrium spp.</i>	24
Table 2.4	Predicted protein spots from World 2D PAGE Repository	26
Table 2.5	Valid spots number detected using PDQuest	29
Table 2.6	Identification of selected proteins from <i>Alexandrium spp.</i>	30
Table 3.1	GTX profiles of <i>A. tamiyavanichii</i> and <i>A. leei</i> at three different phases	44
Table 3.2	STX profiles of <i>A. tamiyavanichii</i> and <i>A. leei</i> at three different phases.	45
Table 3.3	The amount of toxins recorded by both species at different growth phases in molar percentage (mol %)	46
Table 3.4	Total toxins content of <i>A. tamiyavanichii</i> worldwide	50
Table 4.1	Isoelectric focusing steps for 3-10 linear IPG strips	58
Table 4.2	Protein content for each treatment	59

LIST OF FIGURES

<u>Figure No.</u>		<u>Page No.</u>
Figure 2.1	Growth patterns of (A) <i>A. tamiyavanichii</i> and (B) <i>A. leei</i> .	21
Figure 2.2	1DGE and 2DGE images of <i>Alexandrium spp.</i> extracted using three different techniques of cell lysis	23
Figure 2.3	1DGE images of <i>Alexandrium spp.</i> Number 1, 2 and 3 showing bands with difference in intensity between species	25
Figure 2.4	2DGE images of <i>A. tamiyavanichii</i> at different growth phases; (A) exponential, (B) stationary and (C) death	27
Figure 2.5	2DGE images of <i>A. leei</i> at different growth phases; (A) exponential, (B) stationary and (C) death	28
Figure 2.6	Matching set from for spot number 3 between (A) <i>A. tamiyavanichii</i> and (B) <i>A. leei</i> using PDQuest software	34
Figure 2.7	Mechanism reaction of GNATs	35
Figure 2.8	Lipid catabolism by lipases	36
Figure 2.9	Matching set from spot number 5 between (A) <i>A. tamiyavanichii</i> and (B) <i>A. leei</i> using PDQuest software	37
Figure 3.1	Backbone structure of saxitoxin	39
Figure 3.2	Two dimensional structure of voltage-gated sodium channel with binding sites for STX and GTX	40
Figure 3.3	Toxin content of both species recorded at three growth phases of <i>Alexandrium spp.</i> ; exponential, stationary and death	47
Figure 3.4	Mol % of total toxins produced by <i>A. tamiyavanichii</i> (A) and <i>A. leei</i> (B) at three growth phases; exponential, stationary and death	48

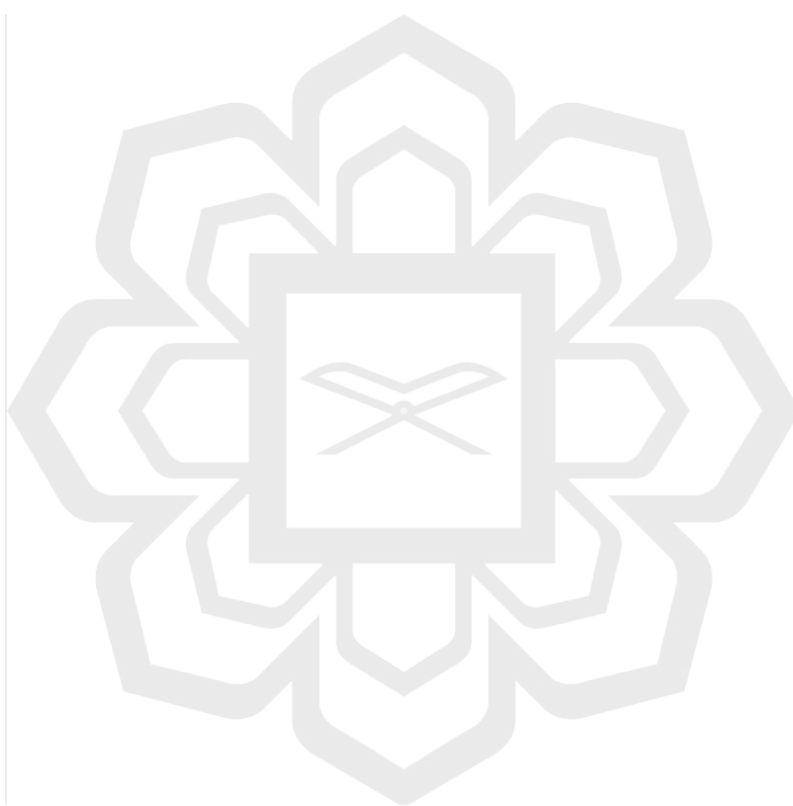
Figure 4.1	1DGE gel image of proteins from <i>C. iridalei</i> which has been exposed with (C1-C2) control, (L1-L3) non-toxic <i>A. leei</i> and (T1-T3) toxic <i>A. tamiyavanichii</i>	60
Figure 4.2	2DGE gel images of proteins from (A) <i>C. iridalei</i> control, which has been exposed with (B) non-toxic <i>A. leei</i> and (C) toxic <i>A. tamiyavanichii</i>	62
Figure 4.3	Images of protein expressed by <i>C. iridalei</i> upon exposure with (A) non-toxic <i>A. leei</i> and (B) toxic <i>A. tamiyavanichii</i> . The yellow circles indicate the unmatched spot present only in the toxic sample	63
Figure 4.4	Histogram view of the two unmatched spots	64



LIST OF ABBREVIATIONS

2D-PAGE	-	Two-Dimensional Polyacrylamide Gel Electrophoresis
2-DE	-	Two dimensional electrophoresis
AK	-	Arginine kinase
APS	-	Ammonium Persulphate
BSA	-	Bovine Serum Albumin
CHAPS	-	3-[(3-cholamidopropyl)dimethylammonio]-1-propanesulfonate
DNA	-	Deoxyribonucleic acid
DTT	-	Dithiothreitol
GTX	-	Gonyautoxin
HAB	-	Harmful algal bloom
HCl	-	Hydrochloric Acid
IEF	-	Isoelectric Focusing
IPG	-	Immobilize pH Gradient
MALDI-TOF	-	Matrix Assisted Laser Desorption Ionization – Time of Flight
PMSF	-	Phenylmethylsulfonyl fluoride
PSP	-	Paralytic shellfish poisoning
RuBisCO	-	Ribulose-1,5-biphosphate carboxylase/oxygenase
SDS	-	Sodium dodecyl sulfate
STX	-	Saxitoxin
tRNA	-	Transfer ribonucleic acid
TEMED	-	Tetramethylethylenediamine

TCA	-	Trichloroacetic acid
Tris	-	Trisaminomethane
UV	-	Ultraviolet



LIST OF SYMBOLS

cm	-	Centimeter
°C	-	degree Celsius
pI	-	Isoelectric Point
<i>g</i>	-	gravitational acceleration
kDa	-	kilodalton
µg	-	microgram
µl	-	Microliter
M	-	Molar
ml	-	Milliliter
nm	-	Nanometer
nmol	-	nanomolar
ppt	-	parts per thousand
%	-	Percent
±	-	Plus-minus
V	-	Volts
v/v	-	volume in volume
w/v	-	weight in volume

CHAPTER ONE

BACKGROUND STUDY

1.1 HARMFUL ALGAL BLOOM (HAB)

Harmful algal bloom (HAB) is a world phenomenon of excessive growth of harmful algae within the water column. Diverse environmental conditions in different regions of the world explain on the tolerance of particular species of microalgae. Enormous fish killings and mortalities of marine mammals in west coast of Florida, severe shellfish poisoning cases within Gulf of Mexico and increasing public health incidents in Alaska are some of the significant impacts of HAB (Kirkpatrick et al., 2010).

Dinoflagellates are one of the primary producers of the aquatic ecosystem. It is also known as causative agent of harmful algal bloom events worldwide apart from diatoms and cyanobacteria. The size for this single-celled dinoflagellate ranges from 2 micrometers to 2 millimetres (Leaw et al., 2010). These microscopic planktonic organisms hold the key to the food chain in aquatic ecosystems. However, the major concern related to the massive growth of this primary producer is due to its capability of synthesizing toxins under different environmental conditions. Blooming of microalgae like dinoflagellates is considered harmful when it brought damaging effects to the biodiversity of the ecosystem. The long term effects of HAB event can only be seen after years of outbreak. For example, the aquaculture industry experienced big loss during harvesting season due to low quality of the fish produced. Besides that, the harmful algae can also cause degeneration of human health as results of consuming contaminated shellfish.

Malaysian coastal waters experienced the aggregation of *Alexandrium spp.* where most occurrences are close to the domiciles and aquaculture area. There were severe cases reported since the past decade for poisoning casualty caused by different types of toxic dinoflagellates such as *Alexandrium minutum* dispersal in Tumpat, Kelantan and *Pyrodinium bahamense* occurrence in Sabah (Lim et al., 2012). Shellfish poisoning related to *Alexandrium spp.* has been occurring in Malaysia since 1990s with the first case of *Alexandrium tamiyavanichii* detected in Sebatu, Malacca followed by the outbreak of *Alexandrium minutum* in Tumpat, Kelantan (Usup et al., 2002; Lim et al., 2006; Lim et al., 2012).

1.2 CATASTROPHIC OUTCOMES OF HAB

There are two major detrimental conditions which prevailed upon blooming of microalgae in Malaysia waters; massive fish killing and shellfish poisonings that can lead to human death. Both toxic and non-toxic microalgae contribute to the damages created out of blooming outbreak.

1.2.1 Enormous Cases of Fish killing

Aquaculture industry is largely affected due to massive fish killings resulted from oxygen depletion and physical infection (Burkholder, Glasgow & Hobbs, 1996). Planktonic dinoflagellates move independently within the water column and commonly assemble at the water surface for maximum UV light exposure. The hypoxia condition induced by excessive growth of microalgae at the water surface leads to the insufficient oxygen supply for fish respiration. Some blooming cases capable of developing 'dead zone' where there is low levels of dissolved oxygen which can cause death to aquatic life.

Massive fish killing also resulted from continuous ingestion on the epidermis tissue of the fish by dinoflagellate species like *Pfiesteria piscicida* (Oikonomou et al., 2012). These dinoflagellates are armored with peduncle that helps their attachment to fish organs like gills and fins where ingestion happens gradually. Unfortunately, this will lead to lethal when toxic microalgae attack speeds up the toxin release upon contact with the ruptured gills (Berge et al., 2012). Blooming of *Cochlodinium polykrikoides* had been reported in Sabah where it caused oxygen depletion and also clogging of the gills due to excessive slime production (Wang et al., 2008; Fukuyo et al., 2011; Harun et al., 2015). The same species also responsible for the huge loss of caged finfish in Penang with minimum loss of 6 million USD recorded (Lim et al., 2014). The excess mucus smothers the gills to a point that suffocates fish in the water column. Fish killing cases of species like barramundi, paddletail snapper, estuary cod and fourfinger threadfin were documented in Tanjung Kupang, Johor which caused economical loss to 250 fishermen and fish farmers (Lim et al., 2014; Razali et al., 2015). The causative agent was identified as *Karlodinium australe*. Economical loss due to mass mortality of fish imposed major alarm to the fisheries industry in Malaysia.

1.2.2 Human Health Deterioration

Shellfish poisoning is another outcome of blooming that indirectly affects human health. The byproducts of toxic dinoflagellates are noxious to marine life as well as human being who consumed shellfish such as clams, mussels and oysters. Shellfish like bivalves and molluscs are the potential vector for dinoflagellates poisoning cases. This is because shellfish feed by straining particulate matters suspended in the water column including dinoflagellates (Ferrante et al., 2013). Filter feeders like shellfish naturally insensible to the toxic dinoflagellates.

The toxins synthesized by dinoflagellates are classified according to its symptoms screened on patients which include paralysis, diarrhea and neurological episodes pertaining to the central nervous system as well as gastrointestinal function (Wang, 2008; Gerssen et al., 2010). There is no specific threshold on the cell abundance of toxic microalgae for the outbreak to be considered harmful. This is because different species vary in terms of toxin synthesis regardless of its cell density. *A. tamiyavanichii* from Japan isolates produced high toxins in low cell count (Kon et al., 2015).

In Borneo Sabah, Paralytic Shellfish Poisoning (PSP) cases have been extensively reported since the first outbreak of *Pyrodinium bahamense* in 1976 (Roy, 1977). Two-hundred and two cases of shellfish poisoning with 7 deaths gave a heads-up to researchers in finding the stimulator of the problems. Meanwhile in Peninsular Malaysia, the most frequent cases of HAB being reported were in Tebrau Straits where the water turbulence is highly influencing the rate of blooming in that area (Lim et al., 2014b). In 1999, Singapore, Changi, Punggol and Sungei Buloh of Tebrau Strait experienced blooming of *Dinophysis caudata* that induces toxicity in mussels but no fatality recorded (Lim et al., 2014b).

1.3 FACTORS INFLUENCING THE OVERGROWTH CELLS

A comprehensive understanding on the physiology and external influences on the targeted species outgrowth has been extensively studied. There are many recurring factors that favor the excessive growth and also toxicity attributes of dinoflagellates which include nutrients enrichment and physical oceanography of the coastal water. However, these factors are interchangeable in accordance with the preferences of the dinoflagellates.

1.3.1 Nutrients Enrichment

Sudden blooming of dinoflagellates is highly affected by its natural conditions where surplus nutrients are deposited. Excessive nutrients loading from land support massive blooming of harmful microalgae along the coastline. The nutrient composition however differs according to species (Guisande et al., 2002; Lim et al., 2010). Untreated water discharged from the industrial effluents, agricultural wastes and domestic sewages may provide excess nutrients that could enhance dinoflagellates to grow excessively in response to the high nutrients input (Anderson, Glibert & Burkholder, 2002). Intensive fish farming technique also contributes towards blooming of dinoflagellates due to excessive usage of feed and chemicals (Chinabut et al., 2006).

1.3.2 Physical Oceanographic Changes

Geographical layouts along the coastline are the major factor that favors the excessive growth of both benthic and planktonic dinoflagellates. The most prevalent element is the water dynamic of the assemblage area as the colonization of the dinoflagellates will disperse upon strong wave action (Pistocchi et al., 2011). This coincides with the fact that most of blooming cases were reported in sheltered area where there is less influence of vertical mixing.

Alexandrium catenella bloom was recorded in Thau Lagoon of Mediterranean Sea where the wave action is restricted hence fostering the growth rate of the microalgae (Vila & Maso, 2005; Anderson, 2007). The lagoon was also holding harbors and jetties that provide full protection of the embayments from tidal actions (Vila & Maso, 2005). Lack of water flowing is one of the conditions that encourage eutrophication to occur.

A study by Hall et al. (2008) claimed that poor mixing of the water column contributed to the stratification of Neuse River Estuary of North Carolina (USA) where dinoflagellate *Karlodinium veneficum* was found dominating the region. All these external circumstances accidentally boost up the cell division where eventually promotes the extensive blooming of dinoflagellates.

1.3.3 Environmental Parameters

Physical variables like water temperature and salinity also support the initiation of blooming and these parameters usually differ according to species. The high temperature is favorable for benthic species and it is further verified when there is no blooming reported during winter (Dagenais-bellefeuille & Morse, 2013). An experimental study by Graneli et al., (2011) concluded that elevated sea water temperature was stimulating the bloom of *Ostreopsis ovata* in Brazil and Italy. The species grew well beyond 26°C with the influence of other promoting factors. A monitoring study in Golden Horn Estuary, Turkey revealed an attack of blooming species *Prorocentrum minimum* within two consecutive years (Taş & Okuş, 2011). Both incidents were recorded during late spring and summer seasons with tremendous increase of cell counts within weeks of occurrence.

Water salinity varied according to places which harbor different types of HAB species. Previous studies had shown variants in terms of growth capabilities and toxic attributes of *Alexandrium spp.* under wide range of salinities. A study conducted by Sukkainen et al. (2013) showed that a wide range salinity changed the toxins level and growth rate of *A. ostenfeldii* and *A. peruvianum* but not their major toxin profile. The study concluded that toxin productions of the two species are strain-specific.

1.4 *Alexandrium* spp. AS PROMINENT CAUSATIVE OF HAB IN MALAYSIA

Dinoflagellate from genus *Alexandrium* predominate West Coast and East Coast of Malaysia with severe occurrences of PSP cases being reported (Fukuyo et al., 2011). *A. tamiyavanichii* was first discovered in Sebatu, Malacca where green mussel breeding project was established. The outbreak of the microalgae resulted in 3 cases of poisoning (Razali et al., 2015). In Tumpat, the agent of toxicity was *A. minutum* with 6 persons hospitalized and one casualty after consuming benthic bivalve (*lokan*) (Razali et al., 2015).

The genomic of *Alexandrium* spp. is highly confined due to its large genome size which then encourages other method of discovering HAB incidents *in situ* (Yang et al., 2010). The studies on protein profiling of dinoflagellates during HABs occurrence is analytical since proteins indirectly control the biochemical process such as cell growth, proliferation and homeostasis of that microorganism (Yang et al., 2010). Besides, the approach can give simultaneous expression of various proteins under one comprehensive proteomic analysis. The significant of this study may lead to a new discovery of proteomic approaches in order to provide prior knowledge for the study of toxicology properties of HABs. Hence, the analysis of the protein as well as toxin level needs to be considered as an important study to understand the biochemical changes in HABs and also the affected shellfish.

1.5 RESEARCH PROBLEM

The cases reported on blooming of toxic-dinoflagellates increases yet few studies that focused on the internal factors that trigger the outbreaks especially in Malaysia (Latib et al., 2015; Danish-Daniel et al., 2017). Proteomic approaches on *Alexandrium sp.* is very potential to understand more on the biochemical activity between toxic and non-toxic species. Few researches were studied on the effects of toxicity on the growth pattern of the dinoflagellates. Therefore this study would highlight on the study of toxicity and growth abilities of *Alexandrium sp.* from Malaysia waters.

1.6 RESEARCH OBJECTIVES

- i. To determine the protein profiles of *A. tamiyavanichii* and *A. leei* at different growth phases
- ii. To identify the function of the proteins expressed in relation to the blooming of *A. tamiyavanichii* and *A. leei*
- iii. To investigate the effect of *Alexandrium spp.* accumulation on protein profile of oyster

CHAPTER TWO

PROTEIN PROFILING AT DIFFERENT GROWTH PHASES OF *A. tamiyavanichii* AND *A. leei*

2.1 INTRODUCTION

2.1.1 Proteomics of Dinoflagellates

Proteomic is a branch of molecular studies of organism where the protein profiling of the selected species will help researchers to understand the biochemical properties of their intact cells. This is relevant as proteins are important molecules that engaged in various cellular activities like homeostasis and cell growth (Wang et al., 2014). This study will be an advantage for researchers in dinoflagellates team to get the best picture of its molecular changes so that further steps to mitigate and control the effects of HABs can be drafted. A list of proteomics involving HAB species was mentioned in a review by Wang et al. (2014). The protein analysis has been conducted in *Alexandrium spp.* worldwide which gave identifications to most proteins expressed within the same genus (Yang et al., 2010; Wang et al., 2013). However, the proteomic of Malaysia isolates is way behind the development as there is no protein profile of local HABs has been established.

2.1.2 Growth Phases of *Alexandrium spp.*

According to Wang et al. (2014), there are four phases of blooming process which include initiation, development, blooming and dissipation of that large microalgae population. These phases apply to most microalgae including dinoflagellates from